



Bruker AV 200 / Topspin 1.3

1. Login and open the program

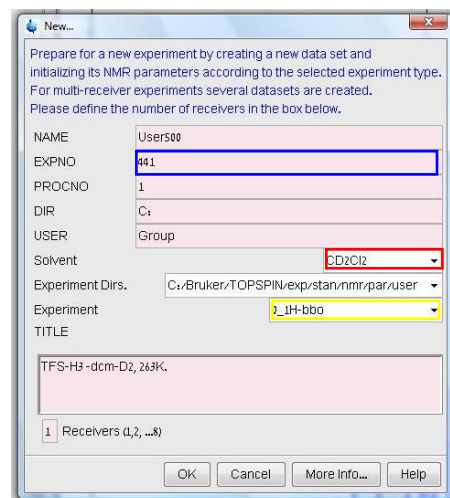
(a) Sign the logbook (name & phone number)

(b) If Topspin is not open, open Topspin. 

2. Create a new file

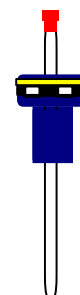
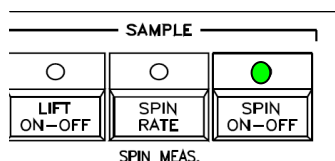
(a) Click on  icon or type: `new/edc`

- **EXPNO** – fill in the “exp number” only integers are allowed.
- **Solvent** – choose your deuterated solvent
- **Experiment** – always begin with **0_1H** choose a set of standard experiments,



3. Insert the sample ([sample preparation](#))

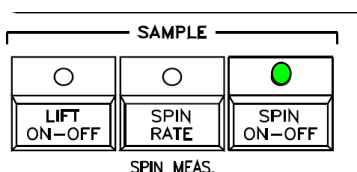
- (a) Make sure the spinner is intact. Place the spinner into the depth measure. Slide the tube down to the bottom (liquid level c.a. ≥ 3 fingers = 600 μ l).
- (b) Hold the tube above the spinner. Wipe the outside of the tube and the spinner.
- (c) BSMS keyboard: press the “**LIFT ON/OFF**” (green = on)



- (d) WAIT until you hear the air flow then set the spinner on the air cushion.
- (e) Press the “**LIFT ON/OFF**” button (no light = off).
- (f) Wait for the green “down” to light up.



4. Spin (possible only for standard tubes)

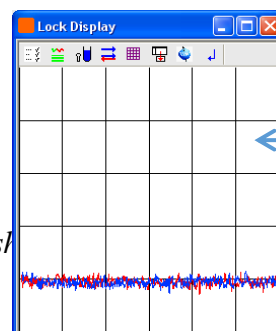
BSMS keyboard:



Press “**SPIN ON/OFF**” (green = on). Wait for steady green light.

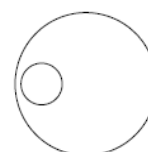
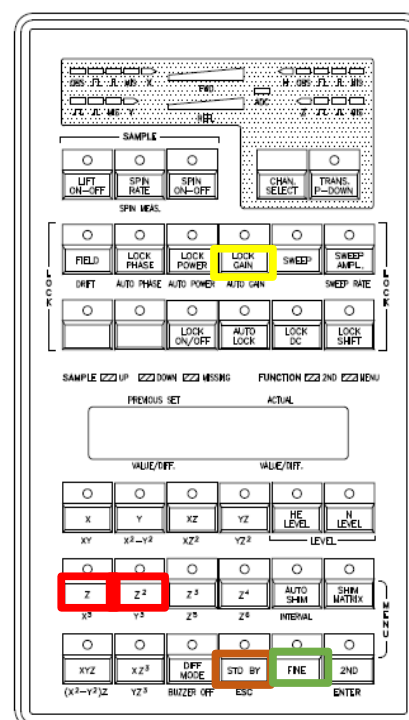
5. Lock

- Click  or type: `lockdisp`.
- Read a shim file by typing: `rsh qnp`.
- Click  or type: `lock`.
- Select your deuterated solvent -
this sets the chemical shift scale. Wait until you get "*lock finish*"



6. Shim

- Press "**LOCK GAIN**" (BSMS board) and use knob to place the locked signal on the second row from the top. Make sure "**FINE**" is on (lit).
- Maximize the signal: Press "**Z**" and move the knob. If signal is improved save the new shim current by pressing "**STD BY**", if not, return to the previous value by pressing "**Z**" again. Repeat for "**Z²**". Repeat again for "**Z**", "**Z²**",... until no further improvement is seen. (If you don't spin your sample shim also "**X**" and "**Y**").
 - if lock signal too high, reduce "**LOCK GAIN**"
 - pressing a lit button will undo changes
 - pressing "**STD BY**" button will save changes
- Press "**STD BY**" when finished.



Control knob

7. Parameter setup

cPars	AcquPars	Title	PulseProg	Peaks
General				
PULPROG =	zg			
TD =	16384			
NS =	1			
DS =	0			
SWH [Hz] =	4006.41			
AQ [s] =	2.0447731			
RG =	11.3			
DW [μ s] =	124.800			
DE [μ s] =	6.00			
D1 [s] =	2.50000000			

(a) Type [ased](#): or click AcquPars tab.

(b) Type [rga](#): wait till you see message "rga finished".

- large **RG** – diluted sample (more scans are required for good S/N)

- small **RG** – concentrated sample

- **RG** =1 too concentrated sample, should be diluted in order to avoid clipping

(c) Click  or type: [zg](#) for preliminary acquisition.

(d) Examine your FID (**time domain**) and your spectrum and choose the right parameters. **avoid clipping of the FID**

Adjust **AQ** (acquisition time) to make sure that the signal vanishes at $\sim 2/3$ of the screen (x-axis). Check that no clipping occurs at y-axis (reduce **RG** if required).


(e) Type: [proc](#) to process you data (FT, phase correction and baseline correction).

Spectrum (frequency domain):

- Adjust "sw" (spectral width in ppm) and "o1p" (center of the spectrum in ppm=resonance frequency) if required.
- Zoom in and check your shimming: if you observe unusual features, splittings or linewidths for ALL your peaks - do more shimming. You can use online shimming by "gs".
- Check again that your **AQ** is sufficient (it might have changed when you adjusted sw).
- Adjust **SI** (ProcPars tab) $\geq 2 * \mathbf{TD}$ to get optimal zero filling.

Set other parameters:



- set **NS** (multiple of 8 for a full phase cycle) and **DS** (=2)
- set **D1** (relaxation delay). Unless you have checked your T_{1s} (relaxation times) and set $AQ + D1 = 5 * T_{1s}(\text{slowest})$ **the spectrum is NOT quantitative!** And the integration is meaningless.

(f) Check the experimental time: click  or type: [expt](#)

8. Acquire a FID

(a) Type [zg](#)

Options during FID acquisition:

- [tr](#)  saves the data and continues acquiring
- [h](#)  HALTs the acquisition and SAVES your FID. *The FID is automatically saved once ns is reached*
- [stop](#) terminates the experiment – data will be lost
- [kill](#) terminates the experiment – data will be lost

9. Processing

Type [proc](#) (performs Fourier Transform ([ft](#)), phase correction ([apk](#)), baseline correction ([abs](#))).

10. Remove your sample

On the BSMS keyboard:

- (a) TURN OFF: " **SPIN ON/OFF**" and " **LOCK ON/OFF** " (no light = off)
- (b) press " **LIFT ON/OFF**" (green = on)
- (c) gently remove your sample from the magnet – lift it up to clear the magnet
- (d) press " **LIFT ON/OFF**" (no light = off)

11. Save your data onto another computer


- (a) Open the nmr200 file icon on the desktop (double-click).
- (b) Click "nmrlab4" network shortcut.
- (c) Drag and drop your files from nmr200 to nmrlab4 and preserve the hierarchy:
<dir>/data/<user>/nmr/<name>/expno

DATA MAY BE DELETED WITHOUT WARNING at any time

Back up your data from **nmrlab4 to your lab computer. Nmrlab4 is not safe and not backed up. Your data is you responsibility!**

12. Open your files on another computer

To open Topspin files on the remote computer:

- (a) Open the program Topspin .
- (b) Navigate to your data using the Browser.
- (c) Drag you data file into the display window.


PLEASE, report problems and bugs to Ira or Shifi (3748)

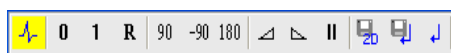
13. More Shimming (on lineshape)


- Shorten AQ and set D1=0sec; set NS=1 DS=0
- Run one scan: `zg` and process `proc`.
- Type: `gs`, choose spectrum view and zoom on solvent (or other narrow) peak.
- BSMS board: Shim slowly (let equilibrate) on **Z**, **Z²** (**X**, **Y** if not spinning) aiming to obtain narrower and higher peak.
- Press “STD BY” when finished.


14. More Processing

- `ft` fourier transform automatically applies the `bc_mod` and `me_mod` specifications
- window functions can optionally be applied to the FID
sensitivity-enhancement ($lb > 0$) `ef`
resolution-enhancement ($lb < 0$, $gb > 0$) `gf`
right-click to alter the display properties
- Automatic phase correction `apk`

Manual phase correction: `.ph` 

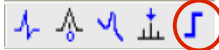


- “0” applies zero-order phase correction
- “1” applies first-order phase correction
- set the cursor with a right-mouse click (“set pivot point” left-click) save and exit: 

- chemical shift calibration `.cal` 


place the cursor at a known position (*e.g.* $\text{CDCl}_3 = 7.26$ ppm)
click the left-mouse button to define its position

- `abs` (automatic baseline correction and automatic integration)

Manual baseline correction: `.basl` 


- highlight the triangle (green) to view differences save and exit.
- For more baseline correction options, type: `bas`

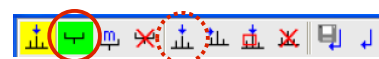
- `ppf` (automatic peak picking)

to toggle display units to relative: click  or type: `.y`

to change parameters for peak picking: `pp`


or type the values in the command line for `mi`, `maxi`, `cy`




Manual/interactive peak picking: `.pp` 



- *define the peak picking range (green box)*
- draw a box around the peaks to pick
- the symbol with the “m” allows you to change the box dimensions
- the peak picking icon allows you to select a single peak with the mouse
- save and exit

- `abs` (automatic integration and baseline correction)
for automatic integration without baseline correction, `absg 0` then `abs`

Manual integration: `.int` 

- to delete all: “select all” with  and delete 
- to define new integrals 
- right-click on an integral to select or calibrate:
- save and exit.

Select / Deselect
Calibrate
Normalize
Lastscal
Delete